

# Biodiversity responses to variation in fire regimes in the coal-seam gas region of south-eastern Queensland

Alan Andersen, Magen Pettit, Teresa Eyre, Garry Cook, Rod Fensham, Belinda Walters & Jodie Hayward

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## Executive summary

Highly fire-prone grassy woodlands dominate the coal-seam gas (CSG) development region of south-eastern Queensland, and the CSG industry has identified potential changes to fire regimes as a priority issue for managing impacts of CSG development on biodiversity. This project addresses the question: *How sensitive is the biota of these woodlands to variation in fire regimes?* It uses results from a controlled fire experiment to model the population dynamics of the dominant eucalypt trees, and examines associations between historical fire regimes and patterns of faunal diversity. Eucalypt populations appear to be limited primarily by water availability, with fire not being a major driver. Changes in tree populations are principally driven by the dynamics of the fire-sensitive native cypress pine (*Callitris columellaris*), which invades the grassy woodlands in the absence of fire.

Patterns of vertebrate diversity in relation to historical fire regimes were examined using 42 survey sites dominated by Poplar box (*Eucalyptus populneus*), located throughout the southern half of the Brigalow Belt bioregion. Information on fire history was obtained from interviews with local land managers, combined with analysis of Landsat imagery over a 25-year period (1987 – 2012). Totals of 48 reptile species, 63 bird species and 6 small mammal species were recorded. Patterns of ant diversity in relation to historical fire regimes were examined using 45 Poplar box/Silver-leaf ironbark (*E. melanophloia*) sites in the Maranoa region, with information on fire history provided by the local land managers. In total, 265 ant species from 50 genera were recorded.

Very frequent fire ( $\geq 3$  fires over 15 yrs) was associated with reduced richness of reptiles generally, and of birds in woodland fragments. The latter appeared to be driven primarily by enhanced abundances of highly aggressive and predatory birds, which have been shown in previous studies to drive out many smaller woodland bird species. For small mammals and ants, richness was lowest in long-unburnt habitat with high cover of native cypress pine. These components of the fauna are adapted to open, grassy habitats, and so habitat favourability for them is reduced by increased canopy cover. We were unable to find any faunal species that was associated with long-unburnt habitat dominated by native cypress pine.

Taken together, our results show that the biota of grassy woodlands in the region is highly resilient to a range of moderate fire frequencies (fires occurring approximately every 10-20 yrs), and that it takes either very high fire frequency (i.e. every few years) or a long-term (several decades) absence of fire to cause substantial change in biodiversity. This situation is very different for fire-sensitive brigalow vegetation, where changed fire regimes due to invasion by buffel grass has been identified as a priority conservation threat. Our findings indicate that any modest change in regional fire regimes is unlikely to have a significant impact on biodiversity in eucalypt-dominated grassy woodlands. However, it is recommended that an ongoing fire monitoring programme be established to ensure that marked changes in regional fire regimes are not occurring as a result of CSG development.

# 1 Introduction

Australia is the most fire-prone of all continents, and its biota is strongly shaped by fire (Bradstock et al. 2012). Appropriate fire regimes are therefore critical for conservation management. Highly fire-prone grassy woodlands dominate the coal-seam gas (CSG) development region of south-eastern Queensland, and the CSG industry has identified fire as a priority issue for managing CSG impacts.

CSG development could potentially influence fire regimes in several ways. On one hand, increased human activity might lead to increased ignition sources and therefore to increased fire frequency. On the other, landscape-scale developments involving long linear structures can restrict the free movement of fire across the landscape, and so reduce fire frequency in isolated patches. The existence of valuable and fire-sensitive infrastructure might also lead to enhanced fire suppression and therefore a managed reduction in the incidence and severity of fires in the landscape. Such changes to the existing fire regime have the potential to lead to significant biodiversity impacts, by altering bio-geochemical cycling and other ecological processes, changing vegetation structure, and promoting invasive species.

The effects of fire on biodiversity have been extensively studied in Australian savannas of far northern Australia. For example, over recent decades four internationally significant long-term fire experiments have been conducted in the Top End of the Northern Territory, at Munmarlary (Russell-Smith et al. 2003), Kapalga (Andersen et al. 2003), the Territory Wildlife Park (Andersen 2014) and Melville Island (Richards et al. 2012), two of which are ongoing. However, no such studies have been conducted in Queensland's CSG region. Fire frequency in this subtropical region is far lower than in the tropical savannas to the north, because the winter dry season is not so severe (Figure 1). Results from the savanna research therefore do not necessarily apply to the grassy biomes of south-eastern Queensland. It is therefore currently not possible to predict the impacts of either increased or reduced fire severity in the CSG region. More generally, the sensitivity of the regional biota to changed fire regimes, and the thresholds at which changed fire regimes cause substantial ecological impact, are unknown.

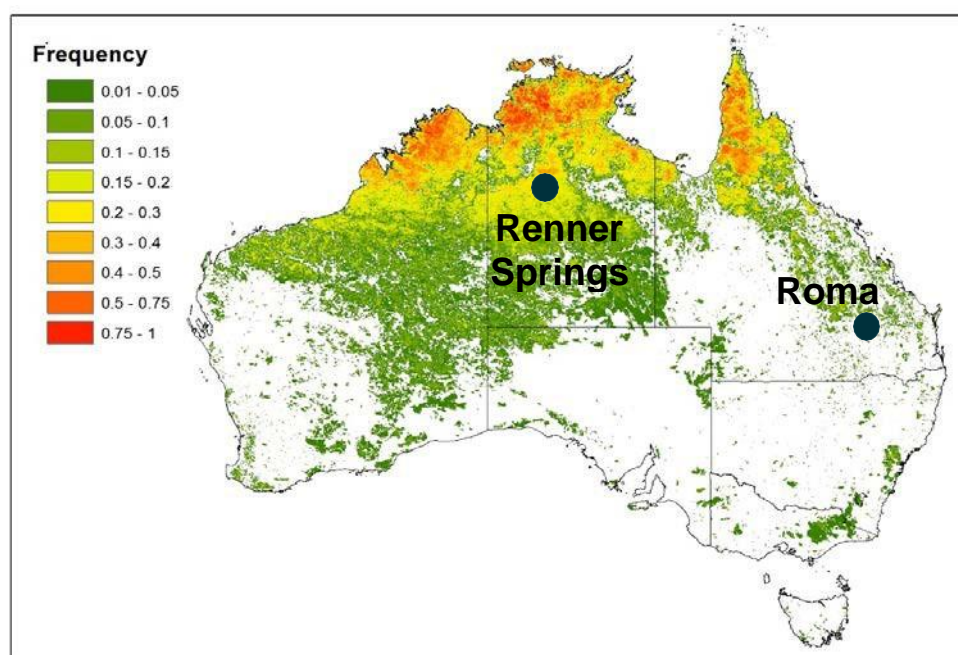
The GISERA fire and biodiversity project addresses this knowledge gap for the regionally dominant and most fire-prone biome, grassy eucalypt-dominated woodlands. It focusses on the question: *How sensitive is the biota of these woodlands to variation in fire regimes?*

The project comprises three components:

1. An experimental study of the effects of fire on eucalypt tree dynamics, led by Garry Cook from CSIRO Land & Water and Dr Rod Fensham from the Queensland Herbarium. It is widely appreciated that in the long-term absence of fire, the grassy woodlands are invaded by the fire-sensitive native cypress pine (*Callitris columellaris*), and that this can cause marked change in vegetation structure (Figure 2). However, the effects of fire on the dominant eucalypt species are poorly known. Results from this component are detailed in the manuscripts: G. D. Cook & R. Fensham. *Tree stand dynamics in central Queensland: Is fire irrelevant?*; and R. Fensham et al. *Climate, not fire, controls woody vegetation*

*dynamics in sub-humid Eucalyptus savanna*. It finds that eucalypt populations are limited primarily by water availability, and that fire is not a major driver. The effects of fire on habitat structure of grassy woodlands in the CSG region are therefore primarily through effects on the density of cypress pine, which increases with time since fire.

2. A study of the relationship between historical fire regimes and current patterns of vertebrate biodiversity, led by Dr Teresa Eyre from the Queensland Herbarium. Results from this component are detailed in the manuscript: Teresa J. Eyre, Daniel J. Ferguson, Luke D. Hogan, Annie L. Kelly, Michael T. Mathieson, Jesse Rowland, Melanie F. Venz, Jian Wang & Alan N. Andersen. *Relative effects of fire on fauna assemblages in fragmented grazing landscapes*. It finds that patterns of vertebrate diversity are strongly associated with historical fire regimes. For reptiles, species richness did not vary among sites experiencing low-to-moderate fire frequency, but was significantly lower at sites experiencing high fire frequency. A similar result was found for birds in woodland fragments, but this was strongly influenced by an interactions between bird species. Frequent fire reduces the shrub density, which provides favourable habitat for predatory birds and the highly aggressive noisy miner, which reduce the density and diversity of small woodland-dependent species. For small mammals, richness was lowest in long-unburnt habitat.
3. A study of the relationship between historical fire regimes and current patterns of ant biodiversity, led by Alan Andersen from CSIRO Land & Water. Ants are a dominant faunal group throughout Australia, and are the most widely used invertebrate indicator group for land management (Andersen & Majer 2004). The responses of ant communities to fire have been extensively studied throughout Australia. Ant diversity in open sclerophyll habitats tends to be promoted by fire, which favours the dominant open-adapted fauna (Andersen 1988; Andersen et al. 2006, 2009; Gosper et al. 2015). Details of this component are provided in the current report, which concludes with a discussion of the management implications of the results of the overall project.





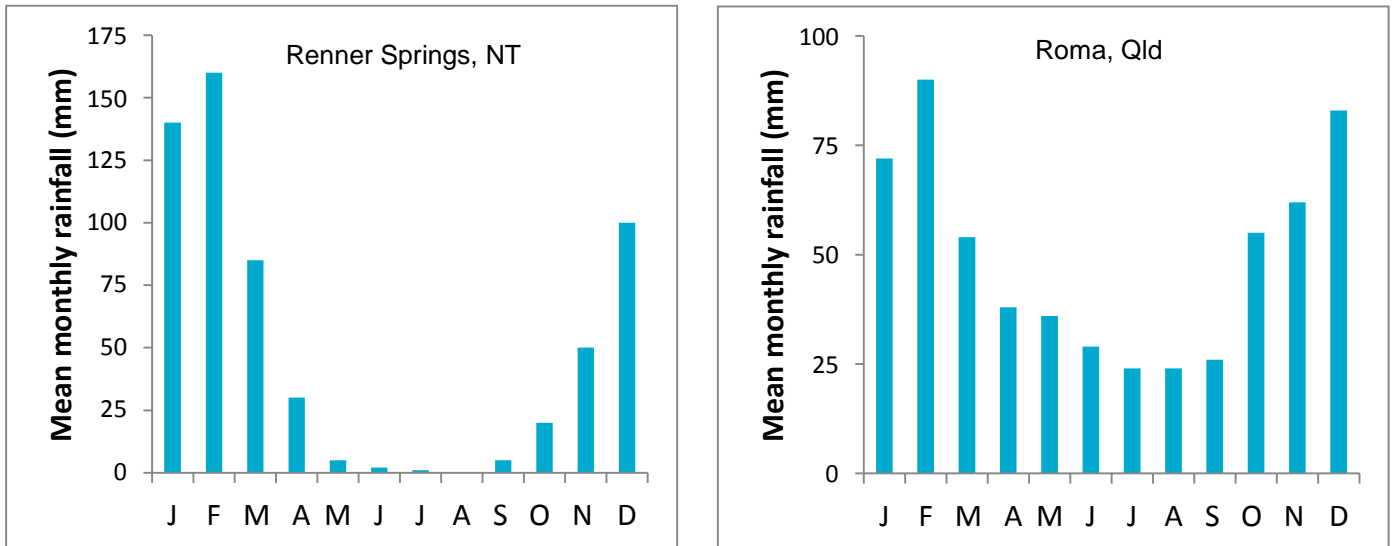


Figure 1. Map showing variation in fire frequency over a 25-yr period across Australia (top). The yellow and red areas (very high fire frequency) denote the tropical savanna zone. Roma (in the CSG region) and Renner Springs have the same mean annual rainfall (600 mm), but fire frequency is far lower in Roma. This can be explained by the more evenly distributed rainfall of Roma (bottom), such that fuels for fire remain relatively moist and therefore less flammable during the dry season.



Figure 2. In the long-term absence of fire, grassy woodlands (left) are invaded by fire-sensitive native cypress pine, resulting in major change to vegetation structure (right). [Photos: A. Andersen]

## 2 Influence of historical fire regimes on ant biodiversity

### 2.1 Methods

#### 2.1.1 Study sites

A total of 45 grassy woodland sites dominated by Poplar box (*Eucalyptus populnea*) and Silver-leaf ironbark (*E. melanophloia*) were studied, distributed across three stations in the Maranoa region: Claravale (14 sites), Currawarra (11) and Myrtleville (20). These stations were chosen because their managers had long associations with them, and had detailed knowledge of their fire histories. The three long-unburnt sites at Myrtleville (M15A, M16A, M17; all with dense cover of cypress pine) were actually located on adjoining Glendonnell Station, within 100 m of its fence and immediately adjacent to frequently burnt sites at Myrtleville. Information on fire history was obtained from the station managers. Fire frequency during the past 30 years ranged from 0 to 12, and time since the last fire ranged from 1 to >50 years (Table 1).

**Table 1. Study sites and their summary fire histories. N – the number of fires in the 30 yrs up to 2013; YRS = the number of years since the last fire, as at 2013.**

CLARAVALE	N	YRS	CURRAWARRA	N	YRS	MYRTLEVILLE	N	YRS
C2B	0	>50	Cu6	3	12	M1	8	1
C3A	3	2	Cu8	3	12	M2	8	1
C3B	3	2	Cu9A	1	12	M3	8	1
C4	3	2	Cu9B	1	12	M4	2	1
C5A	1	15	Cu10	2	4	M5	2	1
C5B	1	5	Cu12	2	1	M6	8	1
C6A	0	>50	Cu13	2	1	M7	8	1
C6B	0	>50	Cu14	1	12	M8	8	1
C7A	1	2	Cu15	2	2	M9	8	1
C7B	1	2	Cu16A	0	>50	M11A	1	3
C8	0	>50	Cu16B	0	>50	M11B	12	1
C9	0	>50				M12	1	3
C10	0	>50				M13	1	3
C11	2	2				M14A	1	3
						M14B	1	3
						M15A	0	>50
						M15B	12	1
						M16A	0	>50

<b>M16B</b>	12	1
<b>M17</b>	0	>50

### 2.1.2 Sampling and sorting

At each site, ants were sampled using a 5 x 4 grid of pitfall traps with 10 m spacing. Traps were partly filled with ethylene glycol as a preservative, and operated for two 48-hr periods, during May and October 2013 at Claravale and Currawarra, and October 2013 and May 2014 at Myrtleville.

Ants from traps were sorted to species, but most could not be named because the great majority of Australian ant species are undescribed. Such species were identified to species group following Andersen (2000), and assigned letter codes (sp. A, etc.) that apply to this study only. The abundance of any species was capped at 50 per trap in order to avoid data distortions caused by extremely high numbers of ants falling into a single trap.

### 2.1.3 Analysis

We analysed variation in mean species richness among sites in relation to fire frequency class and time since last fire using Permanova, considering each station separately. This was also done for the abundances of the ten most common species at each station. Variation in species composition was explored through non-metric multidimensional scaling (NMDS), using Bray-Curtis dissimilarity based on species presence/absence data. Differences among fire frequency classes and times since fire were tested using ANOSIM.

## 2.2 Results

A total of 265 ant species from 50 genera were recorded during the study. The richest genera were *Camponotus* (31 species), *Monomorium* (29), *Pheidole* (25), *Melophorus* (24), *Iridomyrmex* (17), *Meranoplus* (16) and *Tetramorium* (15) (Table 2). The most common species were *Iridomyrmex suchieri* (7.9% total ants in traps), *Rhytidoponera metallica* (6.6%), *Notoncus subdentata* (5.0%), *Iridomyrmex ?chasei* (4.8%), *Monomorium* sp. M (*sordidum* gp.; 4.7%), *Iridomyrmex* sp. C (*rufoniger* gp.; 4.2%), *Monomorium* sp. H (*nigrius* gp.; 4.0%), *Iridomyrmex purpureus* (3.8%), *Monomorium* sp. I (*nigrius* gp.; 3.1%), and *Melophorus* sp. L (*aeneovirens* gp.; 2.1%) (Appendix 1).

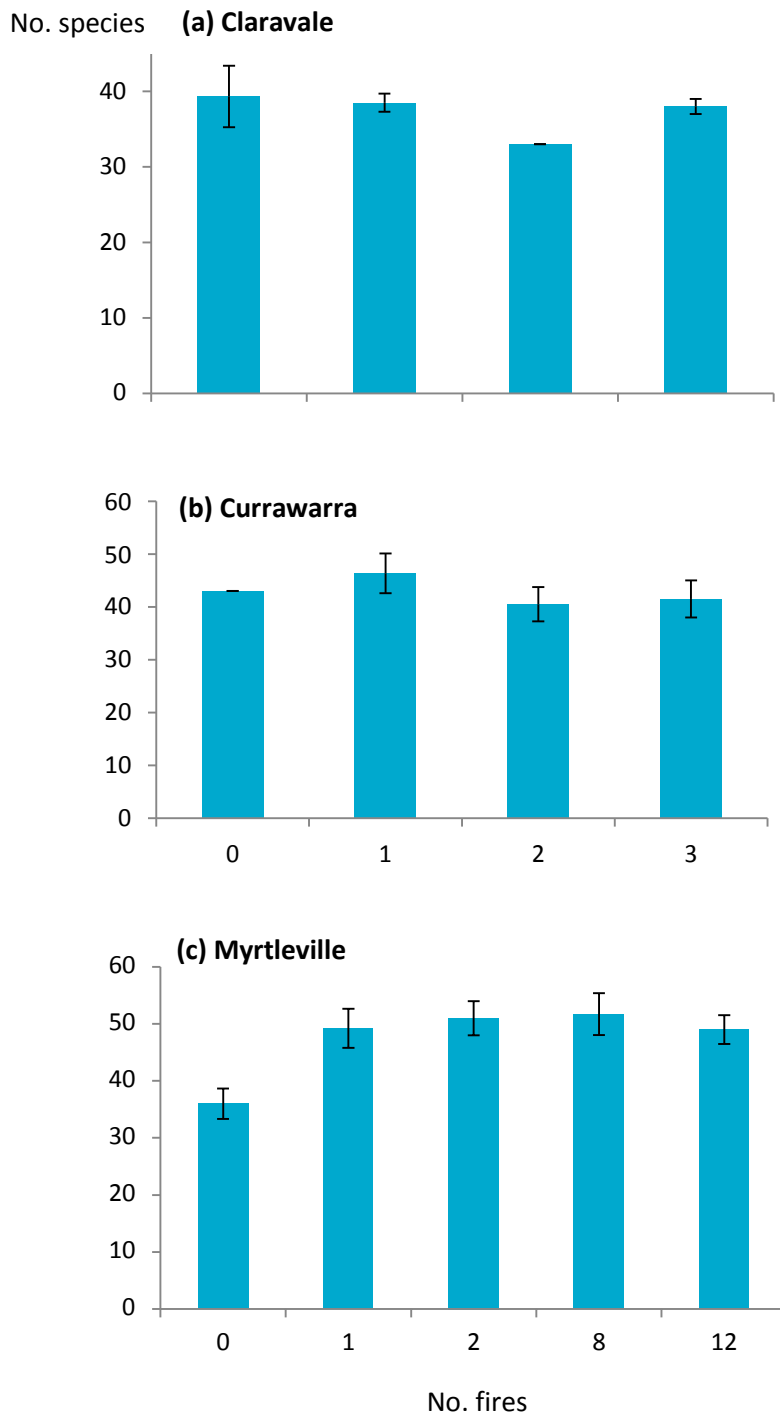
Site species richness ranged from 30 at CL10 to 57 at CL2B (Appendix 1). It averaged 38.4 at Claravale, 42.7 at Currawarra and at 48.3 Myrtleville, with an overall mean of 43.8. At Claravale and Currawarra, site species richness did not vary with either fire frequency class ( $P = 0.887$  and  $0.623$  respectively) or time since last fire ( $P = 0.903$  and  $0.273$  respectively; Figure 3). However at Myrtleville, sites experiencing no fire during the past 30 years had significantly ( $P = 0.004$ ) lower mean richness than at the other sites (Figure 3).

Considering the ten most common species at each site, in only one case did mean abundance vary significantly with fire frequency class: at Myrtleville, *Melophorus* sp. L (*aeneovirens* gp.) was most abundant at moderate to high fire frequencies ( $P = 0.007$ ; Figure 4). There were two cases where mean abundance varied significantly with time since fire: at Claravale, *Monomorium* sp. M (*sordidum* gp.) was more common ( $P = 0.015$ ) at recently (2 yrs) burnt compared with long-unburnt (>50 yrs) sites; and at Currawarra, *Rhytidoponera metallica* was most common ( $P = 0.01$ ) at recently (1 yr) burnt sites.

NMDS revealed only weak (ANOSIM Global R = 0.066) clustering of sites classified according to fire frequency class (Figure 5a). Notably, sites that had remained unburned for >50 years were dispersed throughout ordination space, and were interspersed with sites that had been burnt on 12 occasions during the past 30 years. There was similarly weak (ANOSIM Global R = 0.181) clustering of sites classified according to time since last fire (Figure 5b)

**Table 2. Overview of the ant fauna as recorded in pitfall traps during the study. Figures are numbers of species within genera.**

Subfamily Myrmeciinae		Subfamily Ectatomminae		Subfamily Dolichoderinae	
<i>Myrmecia</i>	1	<b><i>Rhytidoponera</i></b>	5	<i>Anonychomyrma</i>	2
				<i>Arnoldius</i>	1
Subfamily Pseudomyrmecinae		Subfamily Heteroponinae		<i>Dolichoderus</i>	1
<i>Tetraoponera</i>	2	<b><i>Heteroponera</i></b>	1	<i>Iridomyrmex</i>	17
				<i>Leptomyrmex</i>	2
Subfamily Dorylinae		Subfamily Myrmicinae		<i>Ochetellus</i>	3
<i>Aenictus</i>	2	<i>Aphaenogaster</i>	2	<i>Papyrius</i>	1
<i>Cerapachys</i>	5	<i>Cardiocondyla</i>	2	<i>Technomyrmex</i>	1
<i>Sphinctomyrmex</i>	1	<i>Carebara</i>	1		
		<i>Colobostruma</i>	2	Subfamily Formicinae	
Subfamily Amblyoponinae		<i>Crematogaster</i>	6	<i>Acropyga</i>	1
<i>Amblyopone</i>	1	<i>Epopostruma</i>	1	<i>Calomyrmex</i>	2
		<i>Mayriella</i>	1	<i>Camponotus</i>	31
Subfamily Ponerinae		<i>Meranoplus</i>	16	<i>Melophorus</i>	24
<i>Anochetus</i>	1	<i>Mesostruma</i>	1	<i>Notoncus</i>	7
<i>Brachyponera</i>	1	<i>Monomorium</i>	29	<i>Nylanderia</i>	3
<i>Hypoconera</i>	3	<i>Myrmecina</i>	1	<i>Opisthopsis</i>	2
<i>Leptogenys</i>	1	<i>Pheidole</i>	25	<i>Paraparatrechia</i>	7
<i>Odontomachus</i>	2	<i>Podomyrma</i>	3	<i>Polyrhachis</i>	9
<i>Pseudoneoponera</i>	1	<i>Solenopsis</i>	2	<i>Prolasius</i>	1
		<i>Strumigenys</i>	2	<i>Stigmacros</i>	8
		<i>Tetramorium</i>	15		



**Figure 3. Mean ( $\pm$ SE) ant species richness per fire frequency class at each station. There were no significant differences between fire frequency classes at either Claravale or Currawarra (Permanova,  $P > 0.05$ ). At Myrtleville, sites experiencing no fire during the past 30 years had significantly ( $P = 0.004$ ) lower mean richness than at the others.**

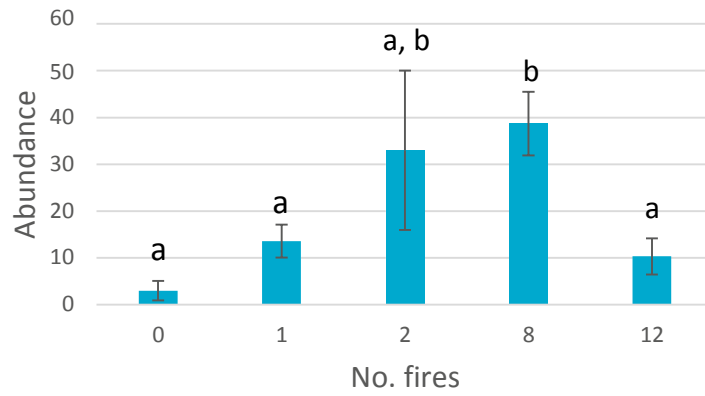


Figure 4. Mean ( $\pm$ SE) abundance of *Melophorus sp. L (aeneovirens gp.)* in relation to fire frequency class at Myrtleville. Fire frequency classes with different letters (a, b) have significantly differences (Permanova,  $P < 0.05$ ) abundances.

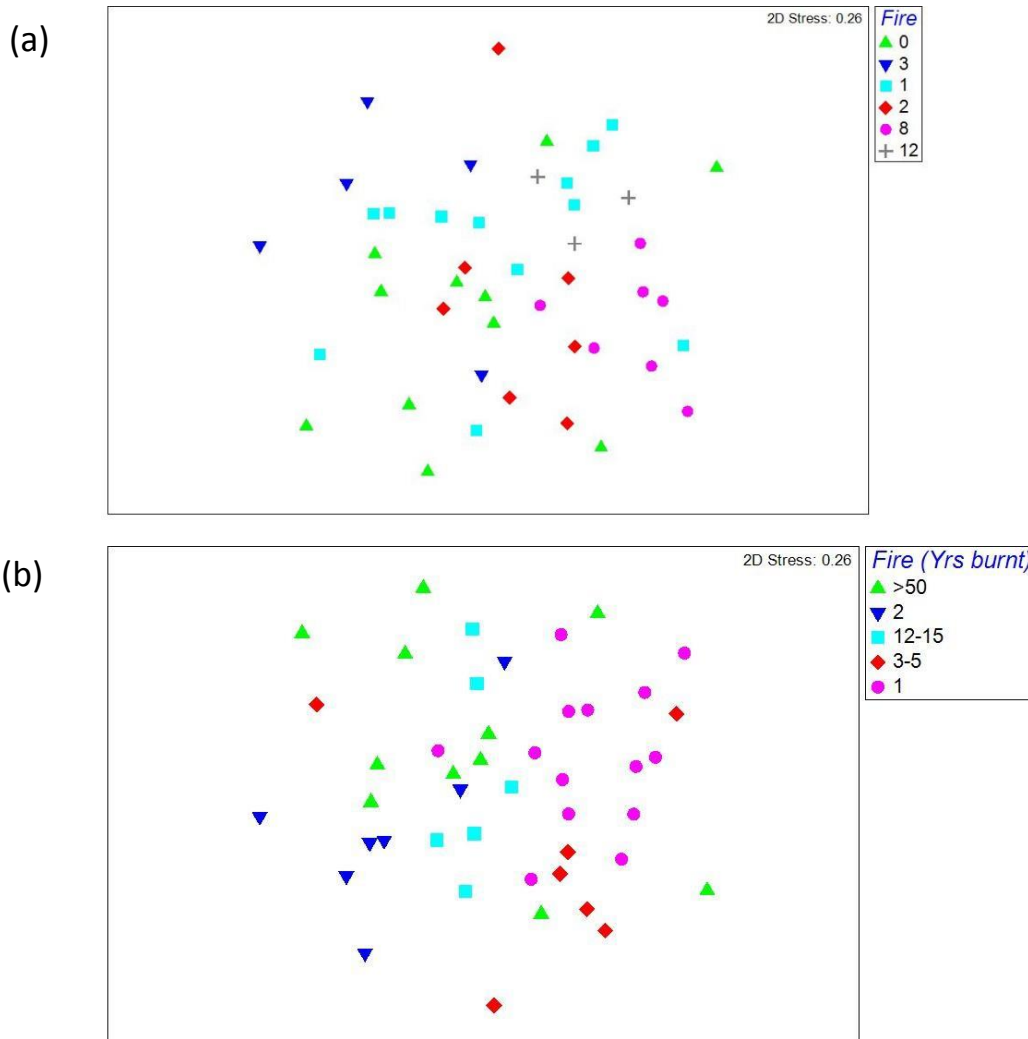


Figure 5. NMDS of study sites classified according to fire frequency class (a; Global  $R = 0.066$ ) and years since last burnt (b; Global  $R = 0.181$ ). Analyses are based on Bray-Curtis dissimilarity, using species presence/absence data.

## 2.3 Discussion

This study represents the first survey of ants in the Maranoa region, and our results show that the Maranoa ant fauna is an extremely diverse one. The fauna is highly noteworthy biogeographically because it has strong representation of taxa characteristic of each of Australia's principle biogeographic realms: Eyrean (arid; e.g. species of *Iridomyrmex*, *Melophorus*, *Meranoplus* and *Tetramorium*); Torresian (tropical; e.g. species of *Anochetus*, *Odontomachus*, *Pseudoneoponera*, *Myrmecina*, *Strumigenys*, *Opisthopsis* and *Calomyrmex*); and Bassian (cool-temperate; e.g. species of *Notoncus*, *Myrmecia*, *Prolasius*, *Stigmacros*, *Epopostruma*, *Colobostruma* and *Mesostruma*). This reflects the location of the Maranoa region at the confluence of these realms. It means that generic diversity is exceptionally high – the 50 genera recorded represent approximately half of Australia's total ant genera (Shattuck 1999), and a far higher proportion of those found outside wet (mostly tropical) forests.

Ant diversity and species composition showed little variation in relation to fire history. Notably, the ant communities of long-unburnt sites were similar to those experiencing high fire frequency. This can be explained by two factors. First, ants suffer very limited mortality during fire because of their below-ground nests, and so the effects of fire on ant communities are primarily indirect, through changes in vegetation structure (Andersen 1988; Andersen et al. 2006). Second, in most cases vegetation structure remained essentially unchanged even in the long-term absence of fire. In the long-term absence of fire, native cypress pine was always present, but typically with low cover, such that the site remained an open grassy woodland (Figure 6). Among our long-unburnt sites, only those at Myrtleville (actually across the fence at Glendonnell) had such a high cover of cypress pine that it transformed vegetation structure (Figure 2). Ant diversity at these sites was substantially lower than at nearby, frequently burnt sites.

We identified three common ant species whose abundances were significantly related to fire history, all of which were most abundant at frequently or recently burnt sites. Two of these – *Melophorus* sp. L (*aeneovirens* gp.) at Myrtleville and *Monomorium* sp. M (*sordidum* gp.) at Claravale, belong to highly thermophilic species groups whose centres of diversity are in the arid zone. They have a particular requirement for open habitats. The third is *Rhytidoponera metallica*, an opportunistic species occurring throughout eastern Australia that is well-known as a disturbance specialist (Hoffmann & Andersen 2003).



**Figure 6. Long-unburnt (>50 yrs) sites at Claravale (CL2B; top) and Currawarra (Cu16B; bottom). Native cypress pine (*Callitris columellaris*) is present, but with low cover, such that the vegetation structure of an open grassy woodland is maintained. [Photos: B. Walters]**



### 3 Conclusion

This project has provided extensive information on the effects of different fire regimes on biodiversity in the grassy, eucalypt-dominated woodlands of the coal-seam gas region of south-eastern Queensland. Modelling of results from a controlled fire experiment in the Desert Uplands indicate that eucalypt populations are limited primarily by water availability, and that fire is not a major driver. Changes in tree populations are therefore driven by the dynamics of the fire-sensitive native cypress pine (*Callitris columellaris*), which invades the grassy woodlands in the absence of fire.

Patterns of faunal diversity in relation to historical fire regimes varied among taxa. Very frequent fire ( $\geq 3$  fires over 15 yrs) was associated with reduced richness of reptiles generally, and of birds in woodland fragments. The latter appeared to be driven primarily by enhanced abundances of highly aggressive and predatory birds, which have been shown in previous studies to drive out many smaller woodland bird species. For small mammals and ants, richness was lowest in long-unburnt habitat with high cover of native cypress pine. These components of the fauna are adapted to open, grassy habitats, and so habitat favourability for them is reduced by increased canopy cover. Notably, we were unable to find any faunal species that was associated with long-unburnt habitat dominated by native cypress pine. This contrasts with biomes in mesic Australia that experience fire at century- rather than decadal scales, where old-growth forest has very high biodiversity value, especially for hollow-nesting fauna (Bradstock et al. 2012).

Taken together, our results show that the biota of grassy woodlands in the CSG region of south-eastern Queensland is highly resilient to a range of moderate fire frequencies (fires occurring approximately every 10-20 yrs), and that it takes either very high fire frequency (i.e. every few years) or a long-term (several decades) absence of fire to cause substantial change in biodiversity. This situation is very different to that for fire-sensitive brigalow vegetation, where increased fire risk due to invasion by the introduced buffel grass has been identified as a priority conservation threat ([http://gisera.org.au/publications/tech\\_reports\\_papers/Brigalow-Belt-PTM-study.pdf](http://gisera.org.au/publications/tech_reports_papers/Brigalow-Belt-PTM-study.pdf)). Our findings indicate that any modest change in regional fire regimes is unlikely to have a significant impact on biodiversity in eucalypt-dominated grassy woodlands. However, it is recommended that an ongoing fire monitoring programme, based on analysis of satellite imagery, be established to ensure that marked changes in regional fire regimes are not occurring as a result of CSG development.

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# Appendix 1

	Claravale											CL10	CL11
	CL2B	CL3A	CL3B	CL4	CL5A	CL5B	CL6A	CL6B	CL7A	CL7B	CL8		
<b>subfamily Myrmecinae</b>													
<i>Myrmecia dimidiata</i>										2	1		3
<i>Myrmecia froggatti</i>													
<i>Myrmecia gilberti</i>						1							
<i>Myrmecia picta</i>								1					
<i>Myrmecia varians</i>		1		2				1		1			
<b>subfamily Pseudomyrmecinae</b>													
<i>Tetraponera punctulata</i>													
<i>Tetraponera</i> nr. <i>punctulata</i>													
<b>subfamily Dorylinae</b>													
<i>Aenictus prolixus</i>													
<i>Aenictus</i> nr. <i>turneri</i>					1								
<i>Cerapachys edentatus</i>								2					
<i>Cerapachys</i> sp. A ( <i>turneri</i> gp.)													
<i>Cerapachys</i> sp. B ( <i>fervidus</i> gp.)													
<i>Cerapachys</i> sp. D ( <i>fervidus</i> gp.)													
<i>Cerapachys</i> sp. E ( <i>brevis</i> gp.)													
<i>Sphinctomyrmex</i> sp. A													
<b>subfamily Ponerinae</b>													
<i>Amblyopone</i> sp. A				1									
<i>Anochetus rectangularis</i>													
<i>Brachyponera lutea</i>	3	4					2		1				
<i>Hypoponera</i> sp. A				1									
<i>Hypoponera</i> sp. B				1									
<i>Hypoponera</i> sp. C													
<i>Leptogenys conigera</i>		1								1	1		
<i>Odontomachus</i> sp. A ( <i>ruficeps</i> gp.)													
<i>Odontomachus</i> sp. B ( <i>ruficeps</i> gp.)													
<i>Pseudoneoponera</i> sp. A ( <i>excavata</i> gp.)	1												
<b>subfamily Ectatomminae</b>													
<i>Rhytidoponera anceps</i>			1	8		1				1			2
<i>Rhytidoponera cristata</i>				3	1		1						1
<i>Rhytidoponera metallica</i>	51	4	15		21	61	4	17	94	8	2	9	10
<i>Rhytidoponera</i> nr. <i>rufescens</i>	12	1				2	1	3		5	3		
<i>Rhytidoponera</i> sp. E ( <i>anceps</i> gp.)													
<b>subfamily Heteroponinae</b>													
<i>Heteroponera imbellis</i>			1										
<b>subfamily Myrmicinae</b>													
<i>Aphaenogaster barbara</i>			51										
<i>Aphaenogaster barbigula</i>							1		6				
<i>Cardiocondyla atalanta</i>					17								
<i>Cardiocondyla nuda</i>	11	1		4		1				7	1	1	49
<i>Carebara</i> sp. A						1							
<i>Colobostruma ellioti</i>			4				1						
<i>Colobostruma</i> sp. B ( <i>alinadis</i> gp.)													
<i>Crematogaster</i> sp. 10 ( <i>australis</i> gp.)	17	54	15		2	1	5	13	5	11	26	2	10
<i>Crematogaster</i> sp. B ( <i>queenslandica</i> gp.)	2			2			1						
<i>Crematogaster</i> sp. C ( <i>queenslandica</i> gp.)	50			75			5	27	12		15		
<i>Crematogaster</i> sp. D ( <i>cornigera</i> gp.)							2				1		
<i>Crematogaster</i> sp. G ( <i>laeviceps</i> gp.)													
<i>Crematogaster</i> sp. J ( <i>laeviceps</i> gp.)													
<i>Epopostruma</i> sp. A	1												
<i>Mayriella spinosior</i>		5	5			1	10	2	3	15			
<i>Meranoplus</i> nr. <i>convexus</i>													
<i>Meranoplus curvispina</i>													
<i>Meranoplus diversoides</i>				7		1			2				
<i>Meranoplus mjobergi</i>													
<i>Meranoplus orientalis</i>													
<i>Meranoplus pubescens</i>	1			5									
<i>Meranoplus similis</i>							1	1					
<i>Meranoplus</i> sp. B ( <i>dimidiatus</i> gp.)		15	1		2						1	2	
<i>Meranoplus</i> sp. I ( <i>dimidiatus</i> gp.)			4										
<i>Meranoplus</i> sp. F ( <i>excavatus</i> gp.)													
<i>Meranoplus</i> sp. N ( <i>excavatus</i> gp.)													
<i>Meranoplus</i> sp. Q ( <i>fenestratus</i> gp.)													
<i>Meranoplus</i> sp. X ( <i>purvi</i> gp.)								4					
<i>Meranoplus</i> sp. A (Group D)								1					
<i>Meranoplus</i> sp. H (Group D)													
<i>Meranoplus</i> sp. L (Group D)													
<i>Mesostruma turneri</i>									1	5			
<i>Monomorium bicorne</i>	1												
<i>Monomorium ?euryadan</i>						1						9	
<i>Monomorium megalaps</i>													
<i>Monomorium rothsteini</i>													15
<i>Monomorium sydneynense</i>				1			8	2	3				
<i>Monomorium</i> sp. A ( <i>carinatum</i> gp.)	1		6		17	2			1	1		1	11
<i>Monomorium</i> sp. R ( <i>carinatum</i> gp.)					2								
<i>Monomorium</i> sp. AA ( <i>carinatum</i> gp.)													
<i>Monomorium</i> sp. AC ( <i>carinatum</i> gp.)	3												
<i>Monomorium</i> sp. AG ( <i>carinatum</i> gp.)								1					
<i>Monomorium</i> sp. B ( <i>castaneum</i> gp.)													1
<i>Monomorium</i> sp. C ( <i>centrale</i> gp.)	4				3		1				31		1
<i>Monomorium</i> sp. D ( <i>eremopilum</i> gp.)							1		7				
<i>Monomorium</i> sp. AH ( <i>eremopilum</i> gp.)								1					
<i>Monomorium</i> sp. BA ( <i>flavipes</i> gp.)													
<i>Monomorium</i> sp. BD ( <i>flavipes</i> gp.)													
<i>Monomorium</i> sp. V ( <i>lacunosum</i> gp.)					1					1			

	CL2B	CL3A	CL3B	CL4	CL5A	CL5B	CL6A	CL6B	CL7A	CL7B	CL8	CL9	CL10	CL11
<i>Monomorium</i> sp. E ( <i>laeve</i> gp.)	15	13	48	37	10		35	16	9	14	1			2
<i>Monomorium</i> sp. G ( <i>laeve</i> gp.)							1				1			
<i>Monomorium</i> sp. BC ( <i>laeve</i> gp.)														
<i>Monomorium</i> sp. H ( <i>nigrus</i> gp.)	32	50	50	16	8	7	82	32	50	6		2	12	
<i>Monomorium</i> sp. I ( <i>nigrus</i> gp.)		10	37		6	3	12	7	15	50			41	5
<i>Monomorium</i> sp. J ( <i>nigrus</i> gp.)					7								1	
<i>Monomorium</i> sp. P ( <i>nigrus</i> gp.)	5									3			2	9
<i>Monomorium</i> sp. Q ( <i>nigrus</i> gp.)														
<i>Monomorium</i> sp. L ( <i>rothsteini</i> gp.)														4
<i>Monomorium</i> sp. M ( <i>sordidum</i> gp.)	5	100	47	100	3		72	13	100	63	2	1	1	51
<i>Monomorium</i> sp. MM ( <i>sordidum</i> gp.)														
<i>Monomorium</i> sp. AF ( <i>sordidum</i> gp.)		10	1											
<i>Myrmecina australis</i>						1								
<i>Pheidole</i> sp. F ( <i>ampla</i> gp.)											19	11	12	
<i>Pheidole</i> sp. K ( <i>ampla</i> gp.)						1								
<i>Pheidole</i> sp. L ( <i>ampla</i> gp.)					9	21								1
<i>Pheidole</i> sp. BD ( <i>ampla</i> gp.)												1		
<i>Pheidole</i> sp. X ( <i>mjobergi</i> gp.)	6	3				1	2	5	3	8		3	3	1
<i>Pheidole</i> sp. G ( <i>mjobergi</i> gp.)														9
<i>Pheidole</i> sp. H ( <i>mjobergi</i> gp.)					15									24
<i>Pheidole</i> sp. O ( <i>pyriformis</i> gp.)			1											
<i>Pheidole</i> sp. P ( <i>pyriformis</i> gp.)					4									
<i>Pheidole</i> sp. W ( <i>pyriformis</i> gp.)		17	1	10										
<i>Pheidole</i> sp. E ( <i>variabilis</i> gp.)				4						4				
<i>Pheidole</i> sp. I ( <i>variabilis</i> gp.)	9				6	17								17
<i>Pheidole</i> sp. J (Group B)	2										13			
<i>Pheidole</i> sp. BC (Group B)														
<i>Pheidole</i> sp. B (Group C)		2			2		10	7	9	2			1	
<i>Pheidole</i> sp. Y (Group C)		1												
<i>Pheidole</i> sp. BG (Group C)					5									
<i>Pheidole</i> sp. A (Group E)	9	20	21	29	9	7	32	7						
<i>Pheidole</i> sp. R (Group E)	3		6					10	30	12	2	3	8	5
<i>Pheidole</i> sp. S (Group E)														
<i>Pheidole</i> sp. U (Group E)							14							
<i>Pheidole</i> sp. UA (Group E)														
<i>Pheidole</i> sp. BBA (Group E)														
<i>Pheidole</i> sp. BBB (Group E)			16											
<i>Pheidole</i> sp. N (Group F)					2								1	
<i>Podomyrma adelaidae</i>														
<i>Podomyrma elongata</i>														
<i>Podomyrma inermis</i>														
<i>Solenopsis</i> sp. A	8	56	31	22			22	21	28	14		1	45	8
<i>Solenopsis</i> sp. B														
<i>Strumigenys</i> sp. A														
<i>Strumigenys</i> sp. B		2	1											
<i>Tetramorium</i> sp. P ( <i>impressum</i> gp.)														
<i>Tetramorium</i> sp. A ( <i>striolatum</i> gp.)			6	1							2			
<i>Tetramorium</i> sp. B ( <i>striolatum</i> gp.)				3		1								
<i>Tetramorium</i> sp. C ( <i>striolatum</i> gp.)			2								1			
<i>Tetramorium</i> sp. CC ( <i>striolatum</i> gp.)														
<i>Tetramorium</i> sp. D ( <i>striolatum</i> gp.)														
<i>Tetramorium</i> sp. F ( <i>striolatum</i> gp.)		19												
<i>Tetramorium</i> sp. G ( <i>striolatum</i> gp.)							1							
<i>Tetramorium</i> sp. H ( <i>striolatum</i> gp.)						1								
<i>Tetramorium</i> sp. I ( <i>striolatum</i> gp.)														
<i>Tetramorium</i> sp. J ( <i>striolatum</i> gp.)														
<i>Tetramorium</i> sp. K ( <i>striolatum</i> gp.)														
<i>Tetramorium</i> sp. L ( <i>striolatum</i> gp.)														
<i>Tetramorium</i> sp. N ( <i>striolatum</i> gp.)														
<i>Tetramorium</i> sp. S ( <i>striolatum</i> gp.)														
<b>subfamily Dolichoderinae</b>														
<i>Anonychomyrma</i> sp. A ( <i>itinerans</i> gp.)														
<i>Anonychomyrma</i> sp. B ( <i>biconvexa</i> gp.)														
<i>Arnoldius</i> sp. A														
<i>Dolichoderus scrobiculatus</i>			3			5	1			2				
<i>Iridomyrmex brunneus</i>	6			4		4	3	6	8		2		2	
<i>Iridomyrmex ?chasei</i>	50	4		100	54		50	50	19				100	
<i>Iridomyrmex discors</i>										4				
<i>Iridomyrmex nr. dromus</i>		1												
<i>Iridomyrmex ?hartmeyeri</i>													3	
<i>Iridomyrmex purpureus</i>	14				13	1				2	65	50		2
<i>Iridomyrmex septentrionalis</i>	100													
<i>Iridomyrmex suchieri</i>	54				60	100	50	51		5	100	17		100
<i>Iridomyrmex suchieroides</i>	7													7
<i>Iridomyrmex nr. suchieroides</i>														
<i>Iridomyrmex</i> sp. A ( <i>anceps</i> gp.)	3												4	
<i>Iridomyrmex</i> sp. G ( <i>bicknelli</i> gp.)	2												1	
<i>Iridomyrmex</i> sp. B ( <i>mjobergi</i> gp.)	1													
<i>Iridomyrmex</i> sp. D ( <i>mjobergi</i> gp.)									9	1				
<i>Iridomyrmex</i> sp. J ( <i>mjobergi</i> gp.)														1
<i>Iridomyrmex</i> sp. P ( <i>mjobergi</i> gp.)					2									
<i>Iridomyrmex</i> sp. C ( <i>rufoniger</i> gp.)		100	100	50					50	8	50			
<i>Leptomyrme rufipes</i>														
<i>Leptomyrme varians</i>														
<i>Ochetellus clarithorax</i>	1	1	1				11			4				
<i>Ochetellus</i> sp. B					1	2		1						
<i>Ochetellus</i> sp. C														
<i>Papyrius</i> sp. A														
<i>Tapinoma</i> sp. A ( <i>minutum</i> gp.)	4			2			1		2	2			1	1
<i>Tapinoma</i> sp. B ( <i>minutum</i> gp.)			35				2	6	8	35				1
<i>Tapinoma</i> sp. C ( <i>minutum</i> gp.)		4					9	11		2				
<i>Technomyrmex antoni</i>	23		2		25		1	4		5		9		3

	CL2B	CL3A	CL3B	CL4	CL5A	CL5B	CL6A	CL6B	CL7A	CL7B	CL8	CL9	CL10	CL11
<b>subfamily Formicinae</b>														
<i>Acropyga</i> sp. A									1					
<i>Calomyrmex albopilosus</i>	3													
<i>Calomyrmex similis</i>	3			1										
<i>Camponotus aeneopilosus</i>	1				3	3	3	2			1		1	2
<i>Camponotus</i> nr. <i>consobrinus</i>														
<i>Camponotus dromas</i>							1		1					
<i>Camponotus ephippium</i>														
<i>Camponotus extensus</i>														
<i>Camponotus loweryi</i>													2	
<i>Camponotus</i> nr. <i>nigriceps</i>	8	2	2	2		1		2	1	1			1	
<i>Camponotus suffusus</i>														
<i>Camponotus</i> nr. <i>vitreus</i>														
<i>Camponotus whitei</i>														
<i>Camponotus</i> sp. B ( <i>claripes</i> gp.)														
<i>Camponotus</i> sp. F ( <i>claripes</i> gp.)								1	1					
<i>Camponotus</i> sp. H ( <i>claripes</i> gp.)	2				1									
<i>Camponotus</i> sp. R ( <i>claripes</i> gp.)				1										
<i>Camponotus</i> sp. W ( <i>claripes</i> gp.)	1		12								2			
<i>Camponotus</i> sp. BA ( <i>claripes</i> gp.)														
<i>Camponotus</i> sp. BC ( <i>claripes</i> gp.)														
<i>Camponotus</i> sp. CA ( <i>claripes</i> gp.)														
<i>Camponotus</i> sp. CB ( <i>claripes</i> gp.)														
<i>Camponotus</i> sp. E ( <i>discors</i> gp.)														
<i>Camponotus</i> sp. BK ( <i>discors</i> gp.)														
<i>Camponotus</i> sp. K ( <i>ephippium</i> gp.)			3											
<i>Camponotus</i> sp. A ( <i>minimus</i> gp.)											1			
<i>Camponotus</i> sp. BF ( <i>nigraeaeus</i> gp.)														
<i>Camponotus</i> sp. BL ( <i>nigraeaeus</i> gp.)														
<i>Camponotus</i> sp. C ( <i>novaehollandiae</i> gp.)														
<i>Camponotus</i> sp. O ( <i>novaehollandiae</i> gp.)														
<i>Camponotus</i> sp. D ( <i>rubiginosus</i> gp.)														
<i>Camponotus</i> sp. G ( <i>rubiginosus</i> gp.)	1		1						2					
<i>Camponotus</i> sp. L ( <i>subnitidus</i> gp.)														
<i>Camponotus</i> sp. X ( <i>subnitidus</i> gp.)														
<i>Melophorus</i> sp. L ( <i>aeneovirens</i> gp.)	1			1	1		4		6	2	2			1
<i>Melophorus</i> sp. BI ( <i>aeneovirens</i> gp.)								1						
<i>Melophorus</i> sp. B ( <i>bruneus</i> gp.)														
<i>Melophorus</i> sp. W ( <i>bruneus</i> gp.)														
<i>Melophorus</i> sp. R ( <i>fieldi</i> gp.)														
<i>Melophorus</i> sp. BH ( <i>fieldi</i> gp.)														
<i>Melophorus</i> sp. A ( <i>froggatti</i> gp.)	6	1	4		3		1					2		1
<i>Melophorus</i> sp. H ( <i>froggatti</i> gp.)		2			3			2	1		2			2
<i>Melophorus</i> sp. C ( <i>mjobergi</i> gp.)				4							1			
<i>Melophorus</i> sp. F ( <i>mjobergi</i> gp.)	6		1		1	2	1					2		1
<i>Melophorus</i> sp. I ( <i>mjobergi</i> gp.)					4					1				
<i>Melophorus</i> sp. K ( <i>mjobergi</i> gp.)	5			3	1						1			7
<i>Melophorus</i> sp. AE ( <i>mjobergi</i> gp.)														
<i>Melophorus</i> sp. AO ( <i>mjobergi</i> gp.)									1	2				
<i>Melophorus</i> sp. BG ( <i>mjobergi</i> gp.)														
<i>Melophorus</i> sp. E ( <i>pillipes</i> gp.)											2	3		
<i>Melophorus</i> sp. AH ( <i>pillipes</i> gp.)														1
<i>Melophorus</i> sp. P ( <i>turneri</i> gp.)														
<i>Melophorus</i> sp. T ( <i>turneri</i> gp.)	1													
<i>Melophorus</i> sp. O ( <i>wheeleri</i> gp.)														
<i>Melophorus</i> sp. S (Group B)														4
<i>Melophorus</i> sp. AT (Group C)														
<i>Melophorus</i> sp. AM (Group J)														
<i>Melophorus</i> sp. CA (Group M)														
<i>Notoncus subdentata</i>	80	27		81	15	3	12	1	34	1	13	5	8	10
<i>Notoncus</i> sp. C ( <i>ectatommoides</i> gp.)														
<i>Notoncus</i> sp. D ( <i>ectatommoides</i> gp.)														
<i>Notoncus</i> sp. E ( <i>enormis</i> gp.)			86				14		1					
<i>Notoncus</i> sp. F ( <i>enormis</i> gp.)	16	3		21		3	6	11		2				
<i>Notoncus</i> sp. G ( <i>enormis</i> gp.)	13				5						9	2	15	
<i>Notoncus</i> sp. I ( <i>giberti</i> gp.)														
<i>Nyländeria rosae</i>	3			50	1	4		1	3	1				3
<i>Nyländeria</i> sp. A ( <i>vaga</i> gp.)			3	51	2		1		2		1	11		2
<i>Nyländeria</i> sp. C ( <i>obscura</i> gp.)		100			5	4								1
<i>Opisthopsis pictus</i>	3			1							1			
<i>Opisthopsis rufithorax</i>	1					1								
<i>Parapatrechina</i> sp. A ( <i>minutula</i> gp.)							1	2		6		1		
<i>Parapatrechina</i> sp. B ( <i>minutula</i> gp.)	3								5	6	1			
<i>Parapatrechina</i> sp. D ( <i>minutula</i> gp.)		8	24	7			1							
<i>Parapatrechina</i> sp. E ( <i>minutula</i> gp.)														
<i>Parapatrechina</i> sp. F ( <i>minutula</i> gp.)														
<i>Parapatrechina</i> sp. G ( <i>minutula</i> gp.)	3													
<i>Parapatrechina</i> sp. H ( <i>minutula</i> gp.)														
<i>Polyrhachis conciliata</i>														
<i>Polyrhachis hookeri</i>						1								
<i>Polyrhachis insularis</i>														
<i>Polyrhachis lata</i>														
<i>Polyrhachis lydiae</i>														
<i>Polyrhachis micans</i>								1						
<i>Polyrhachis prometheus</i>														
<i>Polyrhachis</i> nr. <i>senilis</i>														
<i>Polyrhachis</i> sp. K ( <i>schwiedlandi</i> gp.)														
<i>Prolasius</i> sp. A ( <i>reticulata</i> gp.)														
<i>Stigmacros aciculata</i>	1													
<i>Stigmacros aemula</i>			1											
<i>Stigmacros</i> nr. <i>inermis</i>														
<i>Stigmacros intacta</i>		13										1		
<i>Stigmacros pilosella</i>		1											3	
<i>Stigmacros pusilla</i>														
<i>Stigmacros</i> sp. A ( <i>flavinodis</i> gp.)								1						
<i>Stigmacros</i> sp. N ( <i>pusilla</i> gp.)														
<b>Total</b>	<b>678</b>	<b>656</b>	<b>649</b>	<b>716</b>	<b>348</b>	<b>267</b>	<b>467</b>	<b>354</b>	<b>444</b>	<b>411</b>	<b>332</b>	<b>316</b>	<b>199</b>	<b>372</b>
<b>No. species</b>	<b>57</b>	<b>36</b>	<b>39</b>	<b>39</b>	<b>40</b>	<b>35</b>	<b>35</b>	<b>45</b>	<b>39</b>	<b>40</b>	<b>35</b>	<b>34</b>	<b>30</b>	<b>33</b>



	CU6	CU8	CU9A	CU9B	CU10	CU12	CU13	CU14	CU15	CU16A	CU16B
<i>Monomorium</i> sp. E ( <i>laeve</i> gp.)			16	23				7	8	18	3
<i>Monomorium</i> sp. G ( <i>laeve</i> gp.)											
<i>Monomorium</i> sp. BC ( <i>laeve</i> gp.)			17		4						
<i>Monomorium</i> sp. H ( <i>nigrius</i> gp.)	5	2	54			6		9	50	46	25
<i>Monomorium</i> sp. I ( <i>nigrius</i> gp.)	3		8	5		1	2	12	16	16	10
<i>Monomorium</i> sp. J ( <i>nigrius</i> gp.)		2	50	50	2		4		5		
<i>Monomorium</i> sp. P ( <i>nigrius</i> gp.)			31								
<i>Monomorium</i> sp. Q ( <i>nigrius</i> gp.)											
<i>Monomorium</i> sp. L ( <i>rothsteini</i> gp.)							7				
<i>Monomorium</i> sp. M ( <i>sordidum</i> gp.)	23	2	6	14	68	83	1	51	89	55	35
<i>Monomorium</i> sp. MM ( <i>sordidum</i> gp.)											
<i>Monomorium</i> sp. AF ( <i>sordidum</i> gp.)											
<i>Myrmecina australis</i>											
<i>Pheidale</i> sp. F ( <i>ampla</i> gp.)											
<i>Pheidale</i> sp. K ( <i>ampla</i> gp.)											
<i>Pheidale</i> sp. L ( <i>ampla</i> gp.)											
<i>Pheidale</i> sp. BD ( <i>ampla</i> gp.)											
<i>Pheidale</i> sp. X ( <i>mjobergi</i> gp.)	3	20	3	13	3	1	2			2	11
<i>Pheidale</i> sp. G ( <i>mjobergi</i> gp.)											
<i>Pheidale</i> sp. H ( <i>mjobergi</i> gp.)				4						1	
<i>Pheidale</i> sp. O ( <i>pyriformis</i> gp.)											
<i>Pheidale</i> sp. P ( <i>pyriformis</i> gp.)											
<i>Pheidale</i> sp. W ( <i>pyriformis</i> gp.)	1		1					3			
<i>Pheidale</i> sp. E ( <i>variabilis</i> gp.)											
<i>Pheidale</i> sp. I ( <i>variabilis</i> gp.)		1			18	2		9		6	7
<i>Pheidale</i> sp. J (Group B)		5					3	6			
<i>Pheidale</i> sp. BC (Group B)											
<i>Pheidale</i> sp. B (Group C)				2						15	3
<i>Pheidale</i> sp. Y (Group C)											
<i>Pheidale</i> sp. BG (Group C)											
<i>Pheidale</i> sp. A (Group E)	5	3	3	1	36	34		8	15	12	6
<i>Pheidale</i> sp. R (Group E)		1	1			3			28		1
<i>Pheidale</i> sp. S (Group E)	1										
<i>Pheidale</i> sp. U (Group E)	1			2							
<i>Pheidale</i> sp. UA (Group E)					4						
<i>Pheidale</i> sp. BBA (Group E)		1									
<i>Pheidale</i> sp. BBB (Group E)											
<i>Pheidale</i> sp. N (Group F)											
<i>Podomyrma adelaidae</i>	1		1								
<i>Podomyrma elongata</i>											
<i>Podomyrma inermis</i>											
<i>Solenopsis</i> sp. A	6	3	2	7	4	4	1	1	7	20	10
<i>Solenopsis</i> sp. B									3		
<i>Strumigenys</i> sp. A											
<i>Strumigenys</i> sp. B											
<i>Tetramorium</i> sp. P ( <i>impressum</i> gp.)											
<i>Tetramorium</i> sp. A ( <i>striolatum</i> gp.)	2							1	3		
<i>Tetramorium</i> sp. B ( <i>striolatum</i> gp.)	31	3	1		3	3	6	3	1	1	
<i>Tetramorium</i> sp. C ( <i>striolatum</i> gp.)				1							
<i>Tetramorium</i> sp. CC ( <i>striolatum</i> gp.)				1			2				
<i>Tetramorium</i> sp. D ( <i>striolatum</i> gp.)											
<i>Tetramorium</i> sp. F ( <i>striolatum</i> gp.)											
<i>Tetramorium</i> sp. G ( <i>striolatum</i> gp.)			1			1					
<i>Tetramorium</i> sp. H ( <i>striolatum</i> gp.)										1	
<i>Tetramorium</i> sp. I ( <i>striolatum</i> gp.)											
<i>Tetramorium</i> sp. J ( <i>striolatum</i> gp.)											
<i>Tetramorium</i> sp. K ( <i>striolatum</i> gp.)											
<i>Tetramorium</i> sp. L ( <i>striolatum</i> gp.)											
<i>Tetramorium</i> sp. N ( <i>striolatum</i> gp.)											
<i>Tetramorium</i> sp. S ( <i>striolatum</i> gp.)											
<b>subfamily Dolichoderinae</b>											
<i>Anonychomyrma</i> sp. A ( <i>itinerans</i> gp.)											
<i>Anonychomyrma</i> sp. B ( <i>biconvexa</i> gp.)											
<i>Arnoldius</i> sp. A											
<i>Dolichoderus scrobiculatus</i>			4	6	1	2		2	2		
<i>Iridomyrmex brunneus</i>	12	26	29	30		2	50	20	4	5	4
<i>Iridomyrmex ?chasei</i>	19		35	100	51		100		49	25	8
<i>Iridomyrmex discors</i>			6		27						
<i>Iridomyrmex nr. dromus</i>	2			3	1						
<i>Iridomyrmex ?hartmeyeri</i>											
<i>Iridomyrmex purpureus</i>			53			2			52		
<i>Iridomyrmex septentrionalis</i>						50					
<i>Iridomyrmex suchieri</i>	64	57	50			50	50	2	6	55	
<i>Iridomyrmex suchieroides</i>											
<i>Iridomyrmex nr. suchieroides</i>							50				
<i>Iridomyrmex</i> sp. A ( <i>anceps</i> gp.)						14	50				1
<i>Iridomyrmex</i> sp. G ( <i>bicknelli</i> gp.)			1								
<i>Iridomyrmex</i> sp. B ( <i>mjobergi</i> gp.)											50
<i>Iridomyrmex</i> sp. D ( <i>mjobergi</i> gp.)		1	2							25	14
<i>Iridomyrmex</i> sp. J ( <i>mjobergi</i> gp.)											
<i>Iridomyrmex</i> sp. P ( <i>mjobergi</i> gp.)											
<i>Iridomyrmex</i> sp. C ( <i>rufoniger</i> gp.)		50				100	1			100	50
<i>Leptomyrme rufipes</i>											
<i>Leptomyrme varians</i>			1								
<i>Ochetellus clarithorax</i>	1				1					27	
<i>Ochetellus</i> sp. B										5	
<i>Ochetellus</i> sp. C	4										
<i>Papyrius</i> sp. A											
<i>Tapinoma</i> sp. A ( <i>minutum</i> gp.)	1		1	5				4	11	6	2
<i>Tapinoma</i> sp. B ( <i>minutum</i> gp.)		3							4	7	
<i>Tapinoma</i> sp. C ( <i>minutum</i> gp.)				1		1	12	1	1		2
<i>Technomyrmex antoni</i>		6		54		7		22	24	15	2

	CU6	CU8	CU9A	CU9B	CU10	CU12	CU13	CU14	CU15	CU16A	CU16B
subfamily Formicinae											
<i>Acropyga</i> sp. A											
<i>Calomyrmex albopilosus</i>	1		1	3					18	3	3
<i>Calomyrmex similis</i>											4
<i>Camponotus aeneopilosus</i>		2									
<i>Camponotus</i> nr. <i>consobrinus</i>				1		7				1	
<i>Camponotus dromas</i>								1			
<i>Camponotus ephippium</i>									1		3
<i>Camponotus extensus</i>											
<i>Camponotus loweryi</i>		4	1		1		2		31	4	2
<i>Camponotus</i> nr. <i>nigriceps</i>			1	1	1			6	2	2	
<i>Camponotus suffusus</i>											2
<i>Camponotus</i> nr. <i>vitreus</i>											
<i>Camponotus whitei</i>									1		1
<i>Camponotus</i> sp. B ( <i>claripes</i> gp.)									2		
<i>Camponotus</i> sp. F ( <i>claripes</i> gp.)				1			1		1		
<i>Camponotus</i> sp. H ( <i>claripes</i> gp.)		1							3		
<i>Camponotus</i> sp. R ( <i>claripes</i> gp.)											
<i>Camponotus</i> sp. W ( <i>claripes</i> gp.)											
<i>Camponotus</i> sp. BA ( <i>claripes</i> gp.)											
<i>Camponotus</i> sp. BC ( <i>claripes</i> gp.)											
<i>Camponotus</i> sp. CA ( <i>claripes</i> gp.)											
<i>Camponotus</i> sp. CB ( <i>claripes</i> gp.)											
<i>Camponotus</i> sp. E ( <i>discors</i> gp.)			1				2				
<i>Camponotus</i> sp. BK ( <i>discors</i> gp.)				1							
<i>Camponotus</i> sp. K ( <i>ephippium</i> gp.)											
<i>Camponotus</i> sp. A ( <i>minimus</i> gp.)											
<i>Camponotus</i> sp. BF ( <i>nigraaeneus</i> gp.)											
<i>Camponotus</i> sp. BL ( <i>nigraaeneus</i> gp.)								1	1		
<i>Camponotus</i> sp. C ( <i>novaehollandiae</i> gp.)							1				
<i>Camponotus</i> sp. O ( <i>novaehollandiae</i> gp.)											
<i>Camponotus</i> sp. D ( <i>rubiginosus</i> gp.)								1			
<i>Camponotus</i> sp. G ( <i>rubiginosus</i> gp.)				1					5	1	
<i>Camponotus</i> sp. L ( <i>subnitidus</i> gp.)											
<i>Camponotus</i> sp. X ( <i>subnitidus</i> gp.)											
<i>Melophorus</i> sp. L ( <i>aeneovirens</i> gp.)	3		2	3		1	12	2	6	7	1
<i>Melophorus</i> sp. BI ( <i>aeneovirens</i> gp.)											
<i>Melophorus</i> sp. B ( <i>bruneus</i> gp.)								1			
<i>Melophorus</i> sp. W ( <i>bruneus</i> gp.)											
<i>Melophorus</i> sp. R ( <i>fieldi</i> gp.)								2			
<i>Melophorus</i> sp. BH ( <i>fieldi</i> gp.)											
<i>Melophorus</i> sp. A ( <i>froggatti</i> gp.)		1				5	1		1	5	1
<i>Melophorus</i> sp. H ( <i>froggatti</i> gp.)	3	1		1		3				1	
<i>Melophorus</i> sp. C ( <i>mjobergi</i> gp.)				1							
<i>Melophorus</i> sp. F ( <i>mjobergi</i> gp.)			8				1		1	1	4
<i>Melophorus</i> sp. I ( <i>mjobergi</i> gp.)											
<i>Melophorus</i> sp. K ( <i>mjobergi</i> gp.)	1	6	3	1			2			1	
<i>Melophorus</i> sp. AE ( <i>mjobergi</i> gp.)											
<i>Melophorus</i> sp. AO ( <i>mjobergi</i> gp.)							5	1	4		
<i>Melophorus</i> sp. BG ( <i>mjobergi</i> gp.)											
<i>Melophorus</i> sp. E ( <i>pillipes</i> gp.)		3									1
<i>Melophorus</i> sp. AH ( <i>pillipes</i> gp.)		1	1	22		1		51			
<i>Melophorus</i> sp. P ( <i>turneri</i> gp.)											1
<i>Melophorus</i> sp. T ( <i>turneri</i> gp.)											
<i>Melophorus</i> sp. O ( <i>wheeleri</i> gp.)											2
<i>Melophorus</i> sp. S (Group B)											
<i>Melophorus</i> sp. AT (Group C)		1									
<i>Melophorus</i> sp. AM (Group J)											
<i>Melophorus</i> sp. CA (Group M)											
<i>Notoncus subdentata</i>	82	6	100	59	9	21	8	59	46	100	54
<i>Notoncus</i> sp. C ( <i>ectatommoides</i> gp.)	2						4				
<i>Notoncus</i> sp. D ( <i>ectatommoides</i> gp.)							1				
<i>Notoncus</i> sp. E ( <i>enormis</i> gp.)		3		6							
<i>Notoncus</i> sp. F ( <i>enormis</i> gp.)				2				1	22	1	
<i>Notoncus</i> sp. G ( <i>enormis</i> gp.)		1						3		5	
<i>Notoncus</i> sp. I ( <i>giberti</i> gp.)					1						
<i>Nyländeria rosae</i>	78	1		55	5	8		12			1
<i>Nyländeria</i> sp. A ( <i>vaga</i> gp.)		1	1					11			1
<i>Nyländeria</i> sp. C ( <i>obscura</i> gp.)	31										
<i>Opisthopsis pictus</i>		1	1		1				1	1	
<i>Opisthopsis ruffithorax</i>		1		1		1			4		
<i>Paraparatrechina</i> sp. A ( <i>minutula</i> gp.)			13	4				6			8
<i>Paraparatrechina</i> sp. B ( <i>minutula</i> gp.)				3	12			16			
<i>Paraparatrechina</i> sp. D ( <i>minutula</i> gp.)											
<i>Paraparatrechina</i> sp. E ( <i>minutula</i> gp.)							1				
<i>Paraparatrechina</i> sp. F ( <i>minutula</i> gp.)										2	
<i>Paraparatrechina</i> sp. G ( <i>minutula</i> gp.)											
<i>Paraparatrechina</i> sp. H ( <i>minutula</i> gp.)											
<i>Polyrhachis conciliata</i>											
<i>Polyrhachis hookeri</i>											
<i>Polyrhachis insularis</i>											
<i>Polyrhachis lata</i>					1						
<i>Polyrhachis lydiae</i>		1									
<i>Polyrhachis micans</i>											
<i>Polyrhachis prometheus</i>					2			1			
<i>Polyrhachis</i> nr. <i>senilis</i>	1	1	2	2				1			2
<i>Polyrhachis</i> sp. K ( <i>schwiedlandi</i> gp.)	1										
<i>Prolasius</i> sp. A ( <i>reticulata</i> gp.)				1							
<i>Stigmacros aciculata</i>											
<i>Stigmacros aemula</i>									1		
<i>Stigmacros</i> nr. <i>inermis</i>											
<i>Stigmacros intacta</i>					1						
<i>Stigmacros pilosella</i>								2		1	
<i>Stigmacros pusilla</i>			1								
<i>Stigmacros</i> sp. A ( <i>flavinodis</i> gp.)			1		2	1					
<i>Stigmacros</i> sp. N ( <i>pusilla</i> gp.)											
Total	509	422	662	589	379	499	464	441	694	653	402
No. species	38	45	53	46	36	39	37	40	50	43	43









CONTACT US

t 1300 363 400  
+61 3 9545 2176  
e [csiroenquiries@csiro.au](mailto:csiroenquiries@csiro.au)  
w [www.csiro.au](http://www.csiro.au)

FOR FURTHER INFORMATION

**CSIRO Land & Water**  
Alan Andersen  
t +61 8 8944 8431  
e [Alan.Andersen@csiro.au](mailto:Alan.Andersen@csiro.au)

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